



Bio-refinery Strategy for Fuel Production in Indonesia

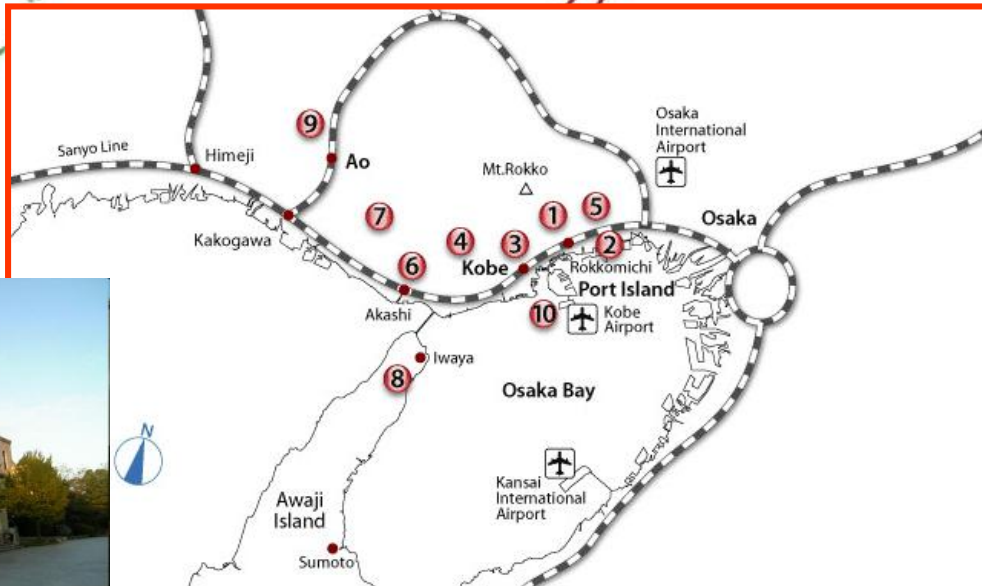
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Kebon Sirih, Jakarta Pusat 10340*

About Kobe and our University



**Kobe University
Bio-production Research Group
(Team Leader: Prof Akihiko Kondo)**



Many Thanks to all member in our Lab.

32 academic staffs including post-doc

43 students including PD candidate

35 technical assistants

Many other peoples

Total 140 (?)

Integrated Biorefinery Research Institute in Kobe University

Only research institute in Japan

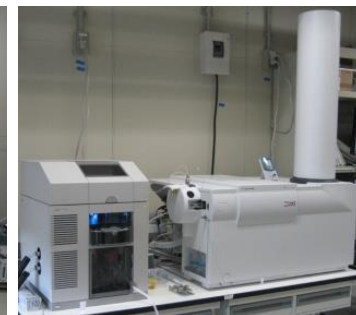
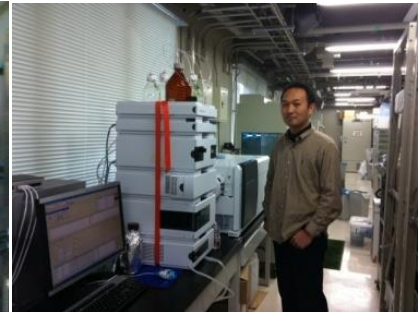
→ Collaborations with more than 20 Japanese Companies.

Research equipment

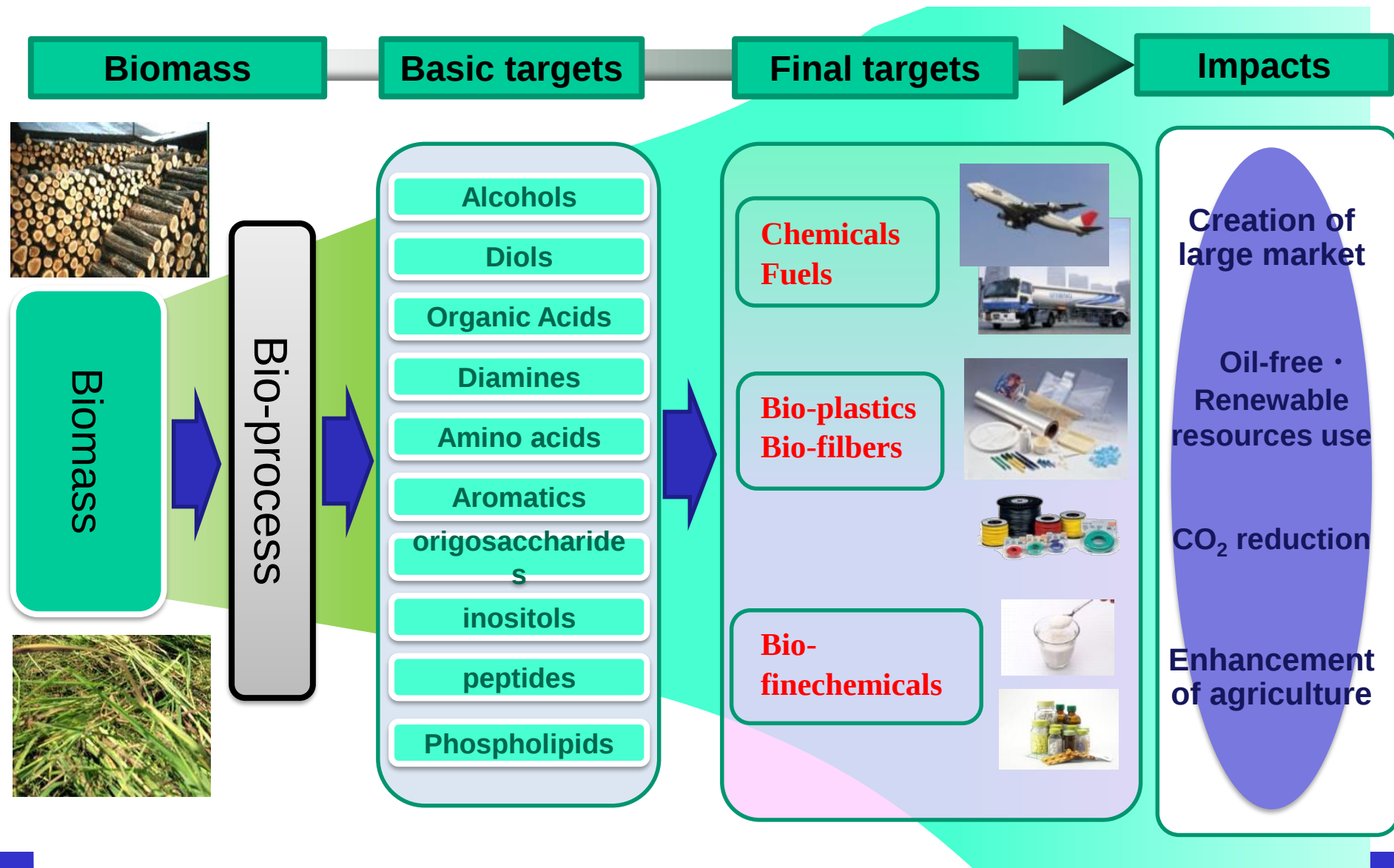
Bioreactor (2L – 100L)
Pretreatment equipment
DNA sequencer
Metabolite analysis equipment

Collaboration with LIPI (2011~)

→ We focused on the utilization possibility of Indonesian's biomass for biorefinery



Systemized bio-refinery process



Utilization possibility of Indonesian's Biomass

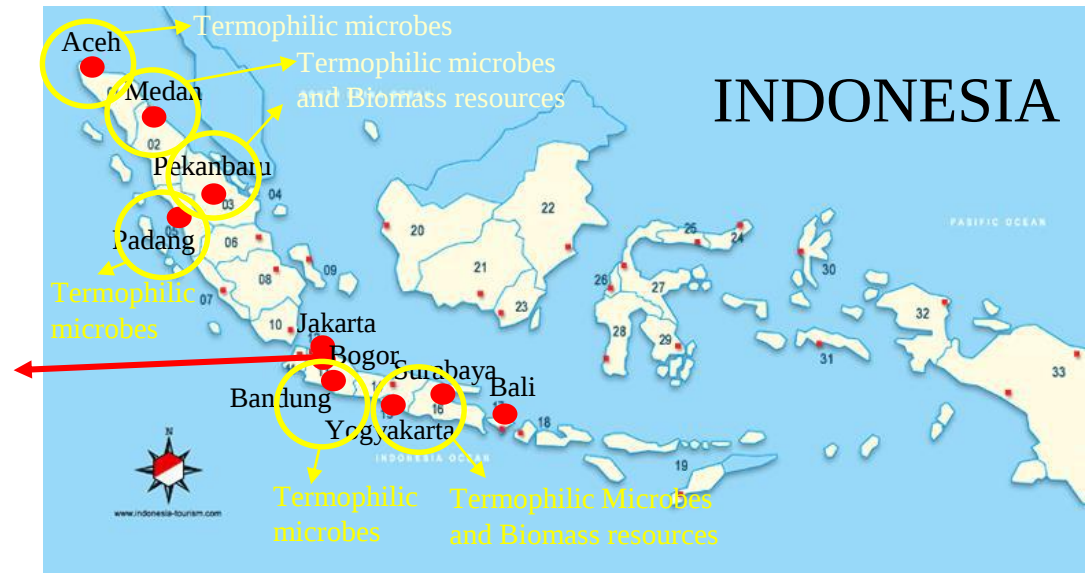
→ Most of waste cellulosic biomass have been burned

Biomass	Generated Place	Productivity [million t/year]	Energy potential [million GJ/year]
Rubber wood (ゴムの木)	Sumatera, Kalimantan, Java	41 (5年間隔で置換)	120
Logging residues (伐木)	Sumatera, Kalimantan	4.5	19
Sawn timber residues (製材残基)	Sumatera, Kalimantan	1.3	13
Plywood and veneer production residues (合板残材)	Kalimantan, Sumatera, Java, Irian Jaya, Maluku	1.5	16
Sugar residues (サトウキビ残渣)	Java, Sumatera, South Kalimantan	Bagasse (バガス): 10 Cane tops: 4 Cane leaves: 9.6	78
Rice residues (稲わら残渣)	Java, Sumatera, Sulawesi, Kalimantan, Bali/Nusa Tenggara	Husk: 12 Bran 2.5 Stalk: 2 Straw: 49	150
Coconut residues (ココナッツ残差)	Sumatera, Java, Sulawesi	Shell: 0.4 Husk: 0.7	7
Palm oil residues (パームオイル残差)	Sumatera new areas: Kalimantan, Sulawesi, Maluku, Nusa Tenggara, Irian Jaya	Empty fruit bunch (EFB): 3.4 Fibres: 3.6 Palm shells (PKC): 1.2	67



→ There are various kinds of biomass for bio-refinery industry

Innovative Bio-Production in Indonesia(iBioI) and Indonesia Culture Collections (InaCC)



iBioI promotes Biomass-based Production of high-valued materials in Indonesia using Indonesia Culture Collection.

InaCC Building in LIPI Cibinong



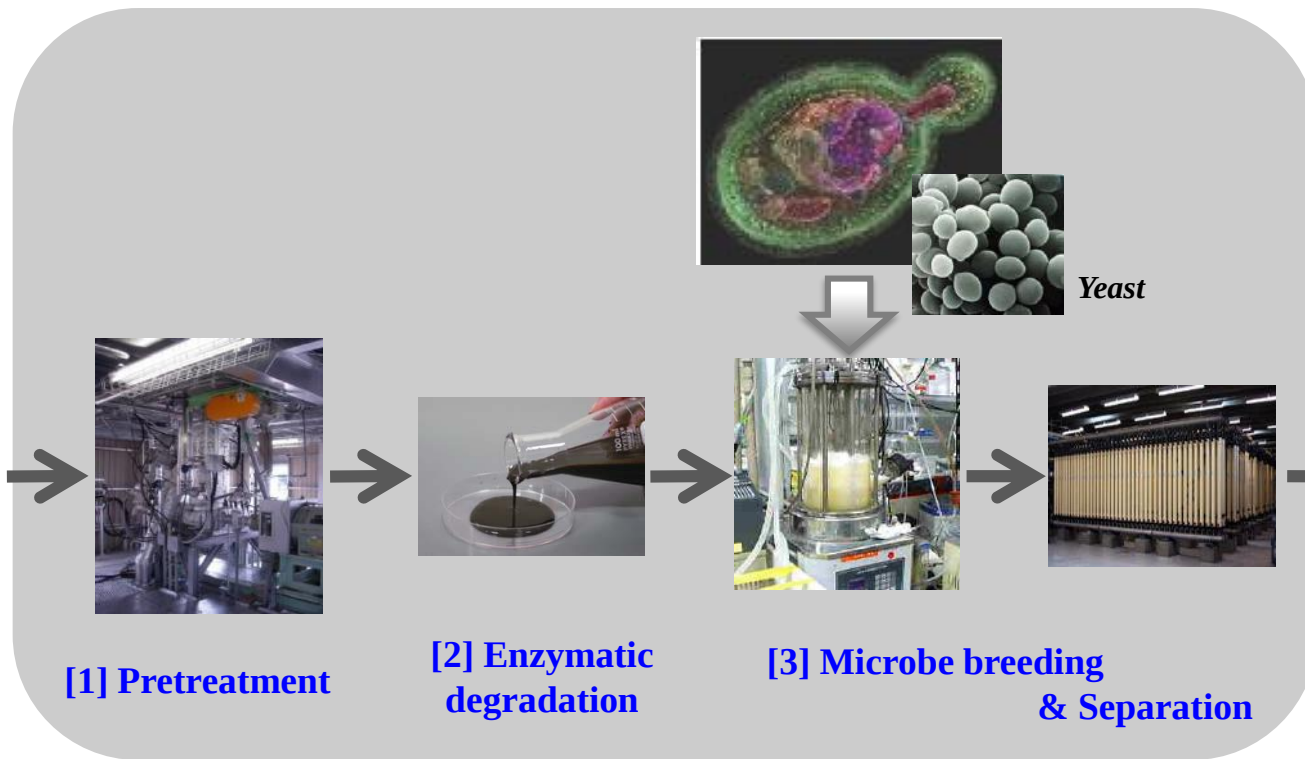
iBioI members



Five Key Technologies in SATREPS project



**Biomass
in Indonesia**

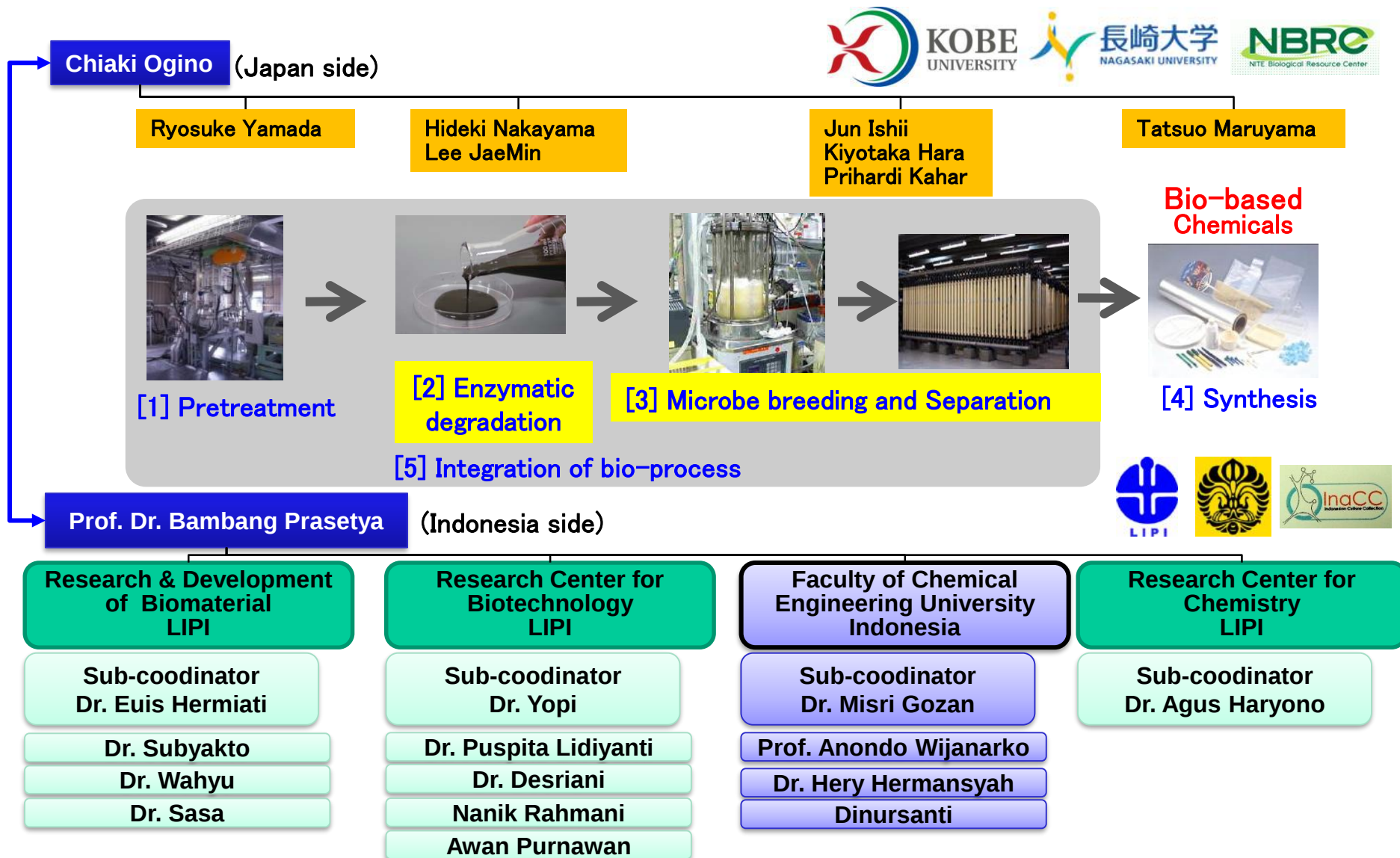


Bio-based

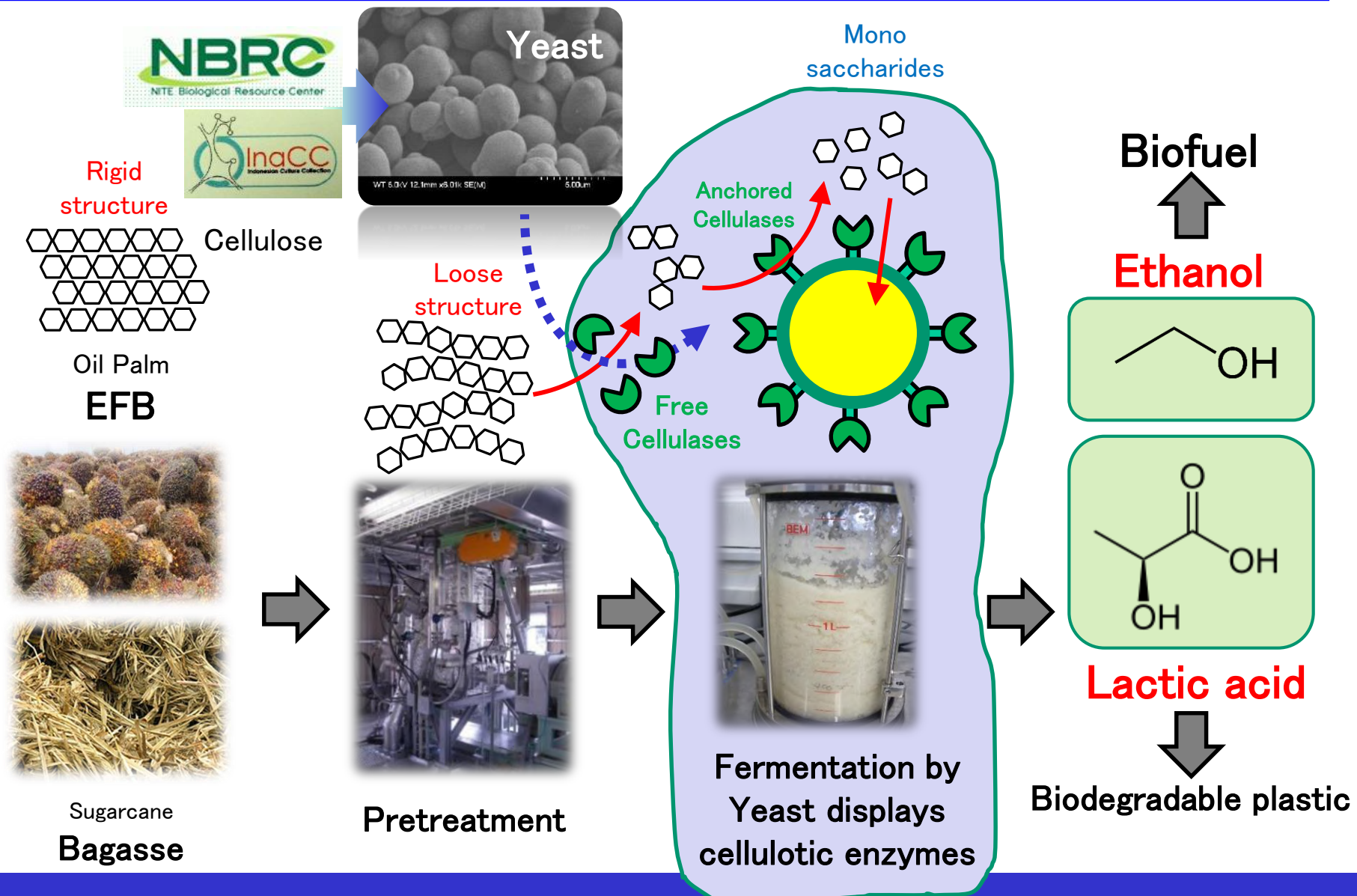


Chemicals

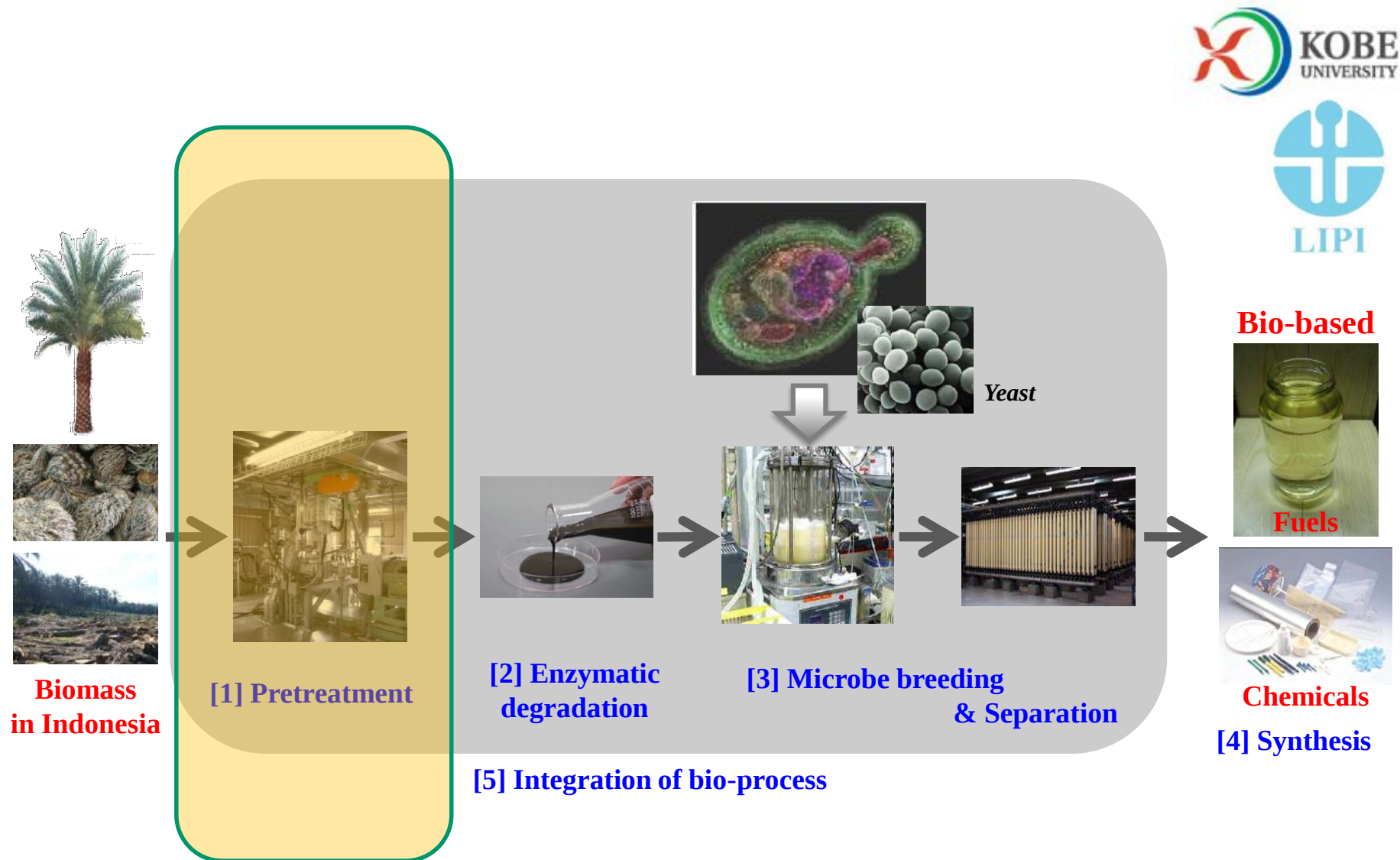
Innovative Bio-production in Indonesia (iBioI) from 2012



Consolidated Conversion of Lignocellulosic Biomass into High-Valued Starting Materials (Ethanol and Lactic Acid)



Five Key Technologies in SATREPS project



Key Cellulosic Constituents

Lignin: 15-25%

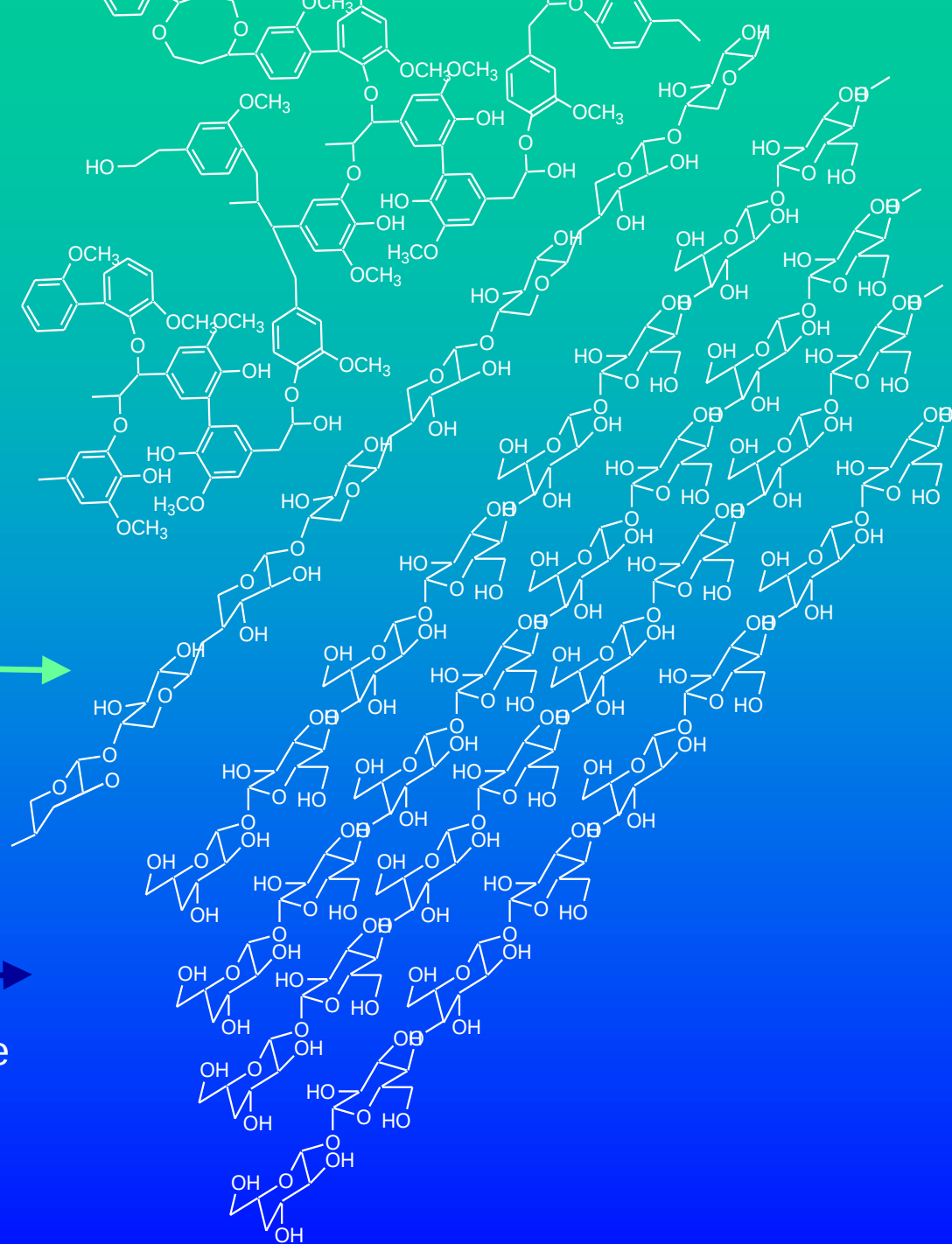
- Complex aromatic structure
- Very high energy content
- Resists biochemical conversion

Hemicellulose: 23-32%

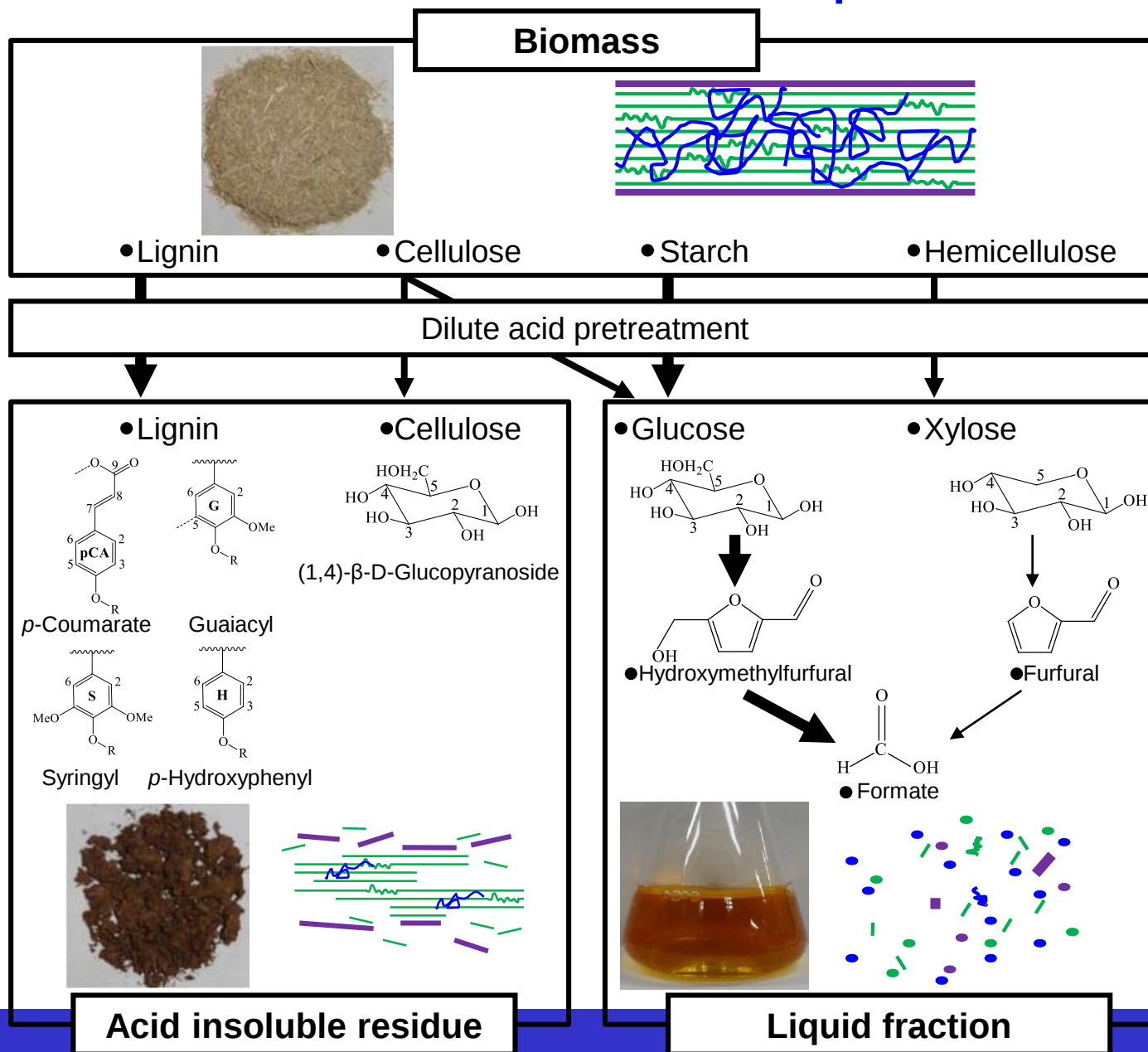
- Xylose is the 2nd most abundant sugar in biosphere
- Polymer of 5C and 6C sugars
- Readily hydrolyzed

Cellulose: 38-50%

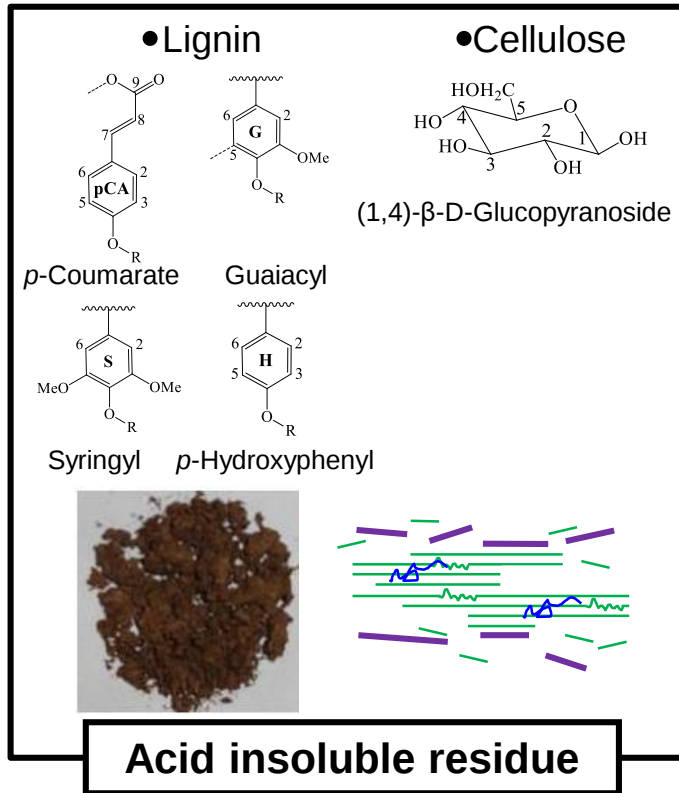
- Most abundant form of C in biosphere
- Polymer of glucose
- Resistant to hydrolysis



Material follow of dilute acid pretreatment



Problem of dilute acid pretreatment



Dilute acid pretreatment can not separate cellulose and lignin.

Lignin inhibit enzymatic hydrolysis of cellulose.



Organosolv pretreatment can separate cellulose and lignin. But, this method require high amount of organic solvent.

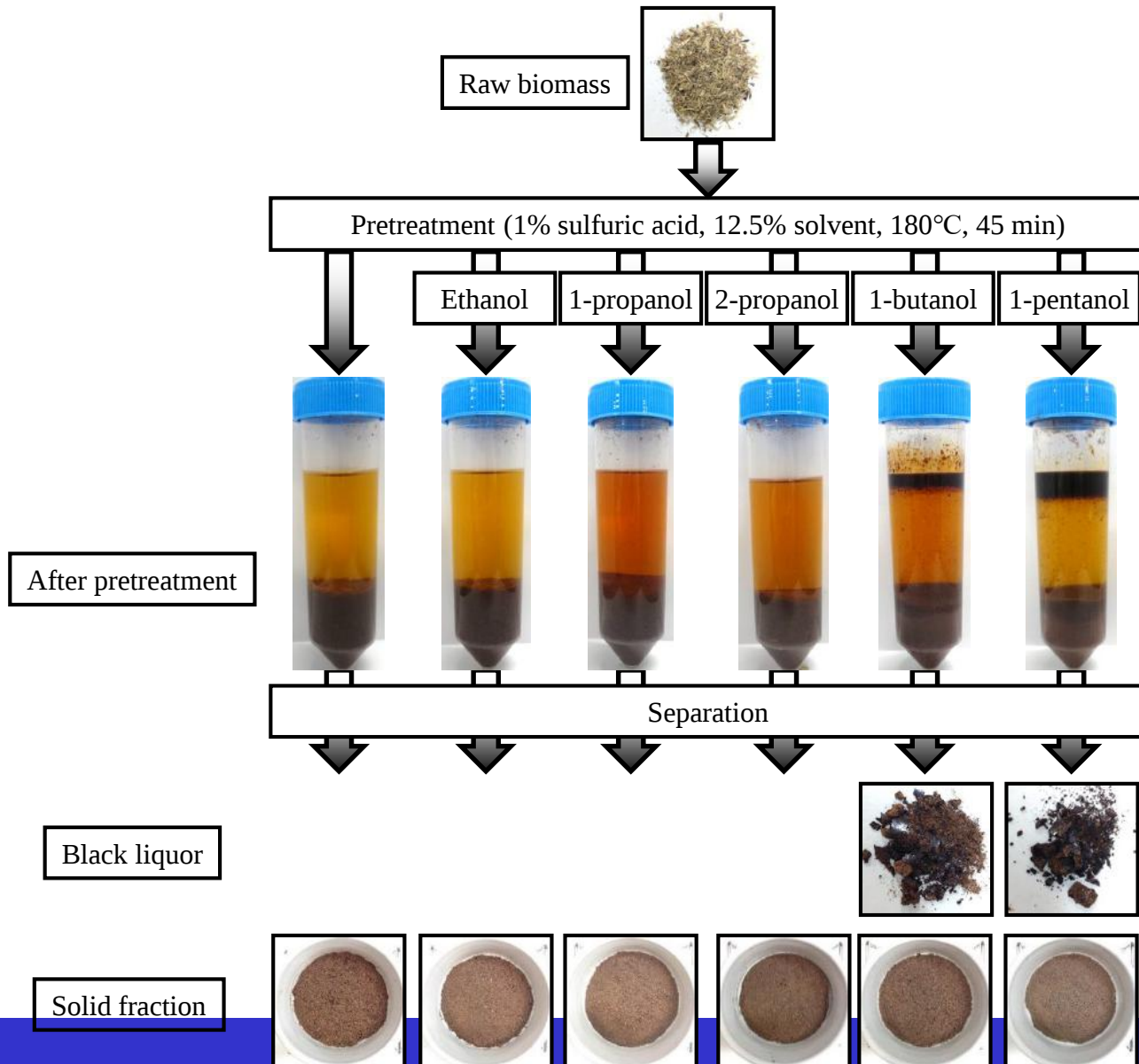
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High cost.

Decreasing the concentration of organic solvent is necessary to reduce the pretreatment cost.

Which of the organic solvent is suitable for pre-treatment?

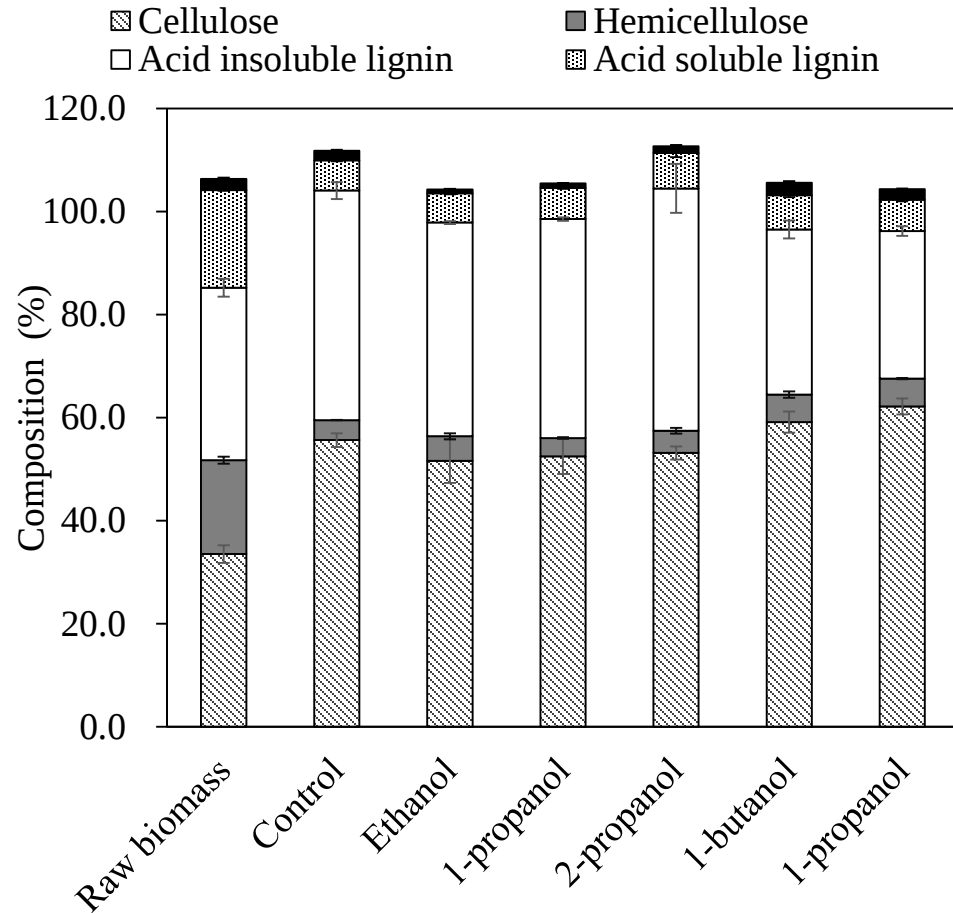
Influence of solvent type on organosolv fractionation of sorghum bagasse



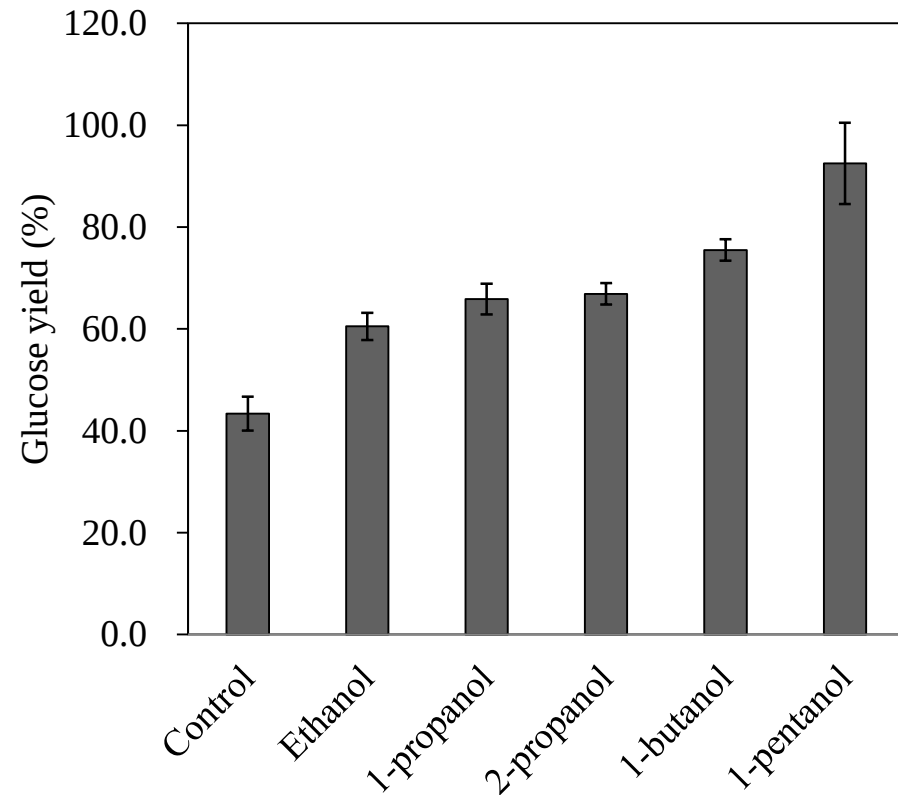
Three fractions (solid, liquid, and black liquor) were obtained with 1-butanol or 1-pentanol as the solvent.

Analysis of Cellulose-enriched solid fraction

Composition of solid fraction

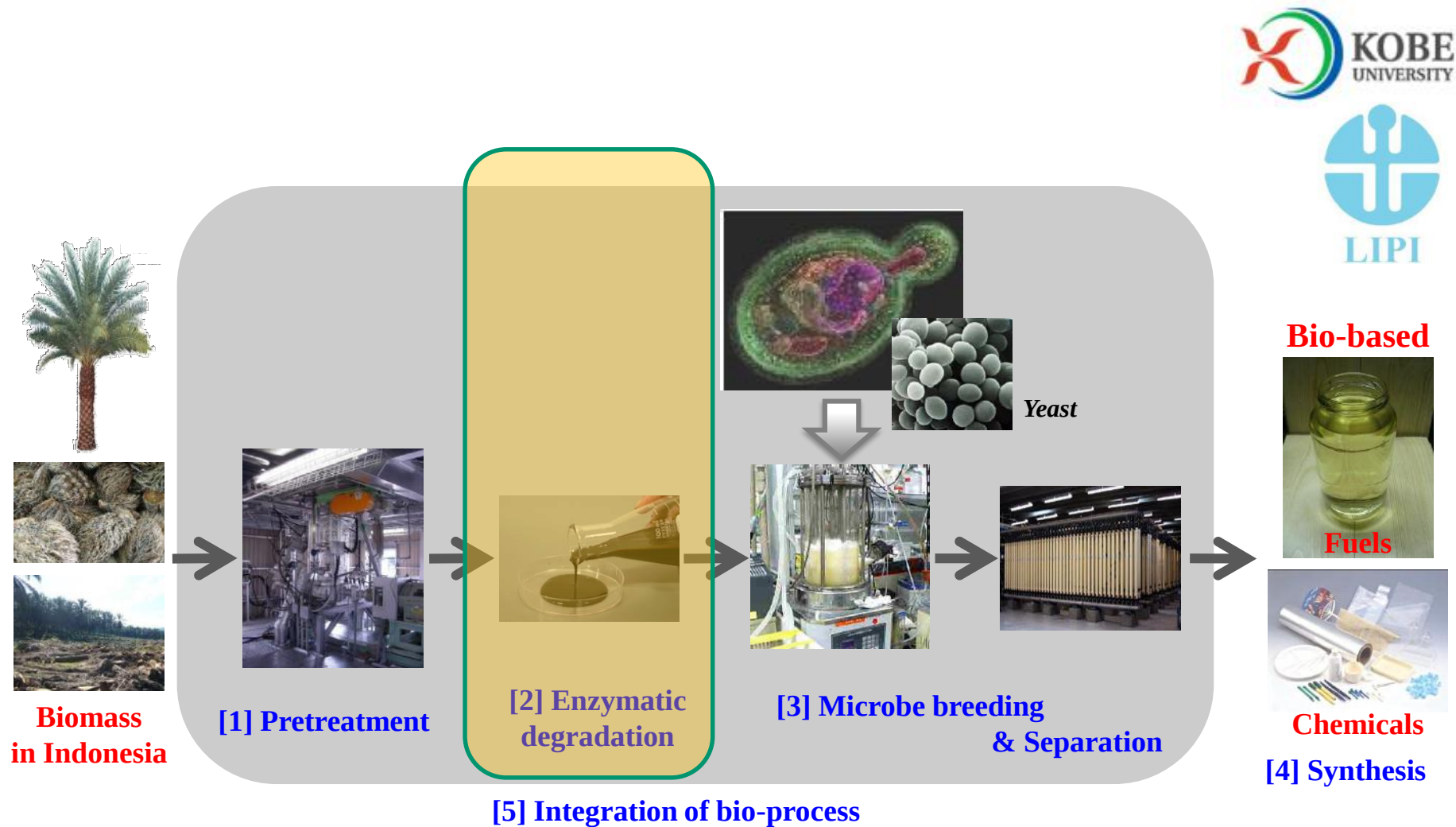


Enzymatic saccharification (6.6FPU/g biomass)



Lignin removal from solid fraction was higher when 1-butanol or 1-pentanol was used as the solvent. And, lignin removal enhance the enzymatic saccharification efficiency.

Five Key Technologies in SATREPS project



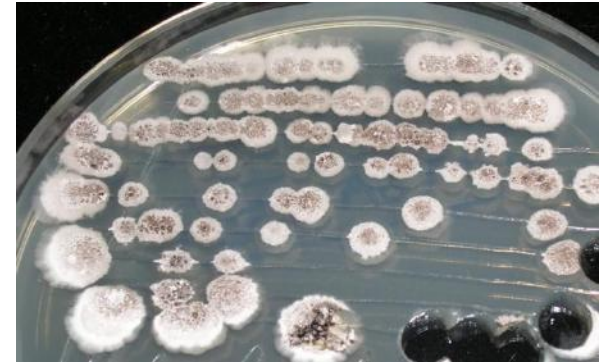
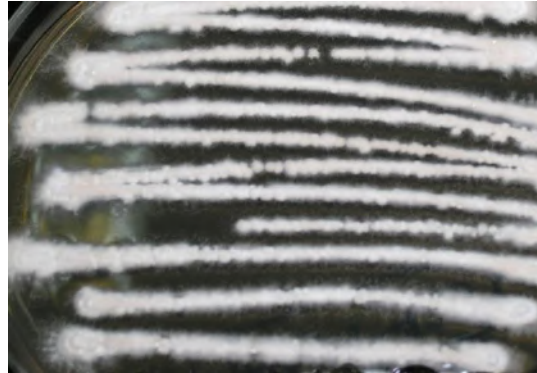
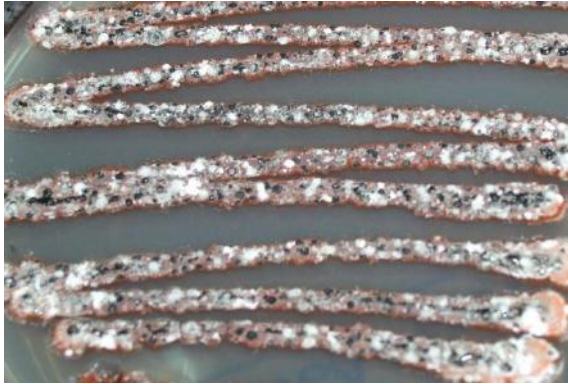
The sources of Actinomyces Isolates in Indonesia

Sampling sites in Indonesia

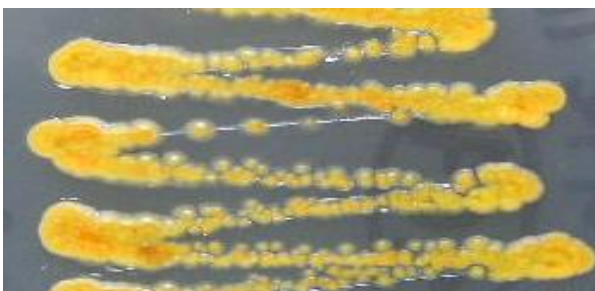
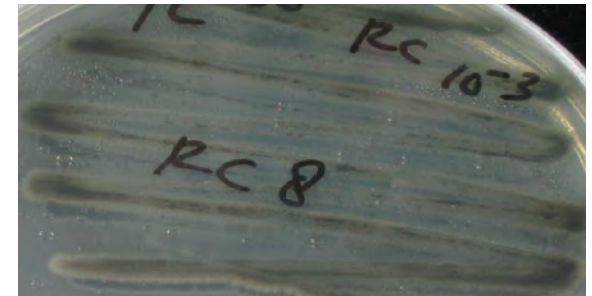
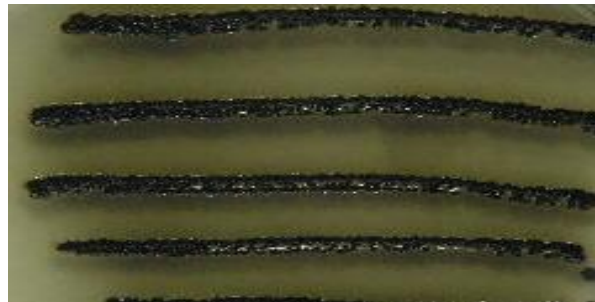


The kinds of Actinomyces Isolates in Indonesia

The genus *Streptomyces*, *Kitasatospora*, *Streptacidiphilus*



The genus *Actinoplanes*, *Micromonospora*, *Nocardia*, *Gordonia*, *Dermatophilus*

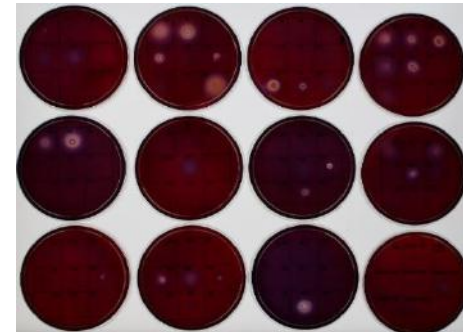


Screening of Indonesia Rare Actinomycetes Isolates based on Halo-formation and Congo Red Analysis

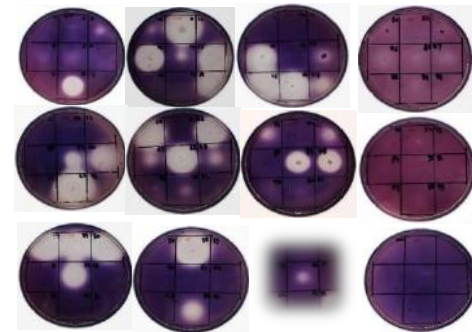
No	Isolate code	Name of isolates
1	ID.04-361	<i>Streptomyces fumigatiscleroticus</i>
2	ID.04-364	<i>Streptomyces griseovariabilis</i>
3	ID.04-372	<i>Actinomadura citrea</i>
4	ID.04-454	<i>Streptomyces turgidiscabies</i>
5	ID.04-455	<i>Kitasatospora putterlickiae</i>
6	ID.04-487	<i>Streptomyces albulus</i>
7	ID.04-502	<i>Streptomyces platensis</i>
8	ID.04-509	<i>Kitasatospora recifensis</i>
9	ID.04-523	<i>Streptomyces galbus</i>
10	ID.04-532	
11	ID.04-539	<i>Streptomyces lipmanii</i>
12	ID.04-540	<i>Kitasatospora kepongensis</i>
13	ID.04-542	<i>Streptomyces glaucescens</i>
14	ID.04-544	<i>Streptomyces galbus</i>
15	ID.04-561	
16	ID.04-570	
17	ID.04-583	
18	ID.04-590	<i>Streptomyces argillaceus</i>
19	ID.04-651	<i>Streptomyces maritimus</i>
20	ID.04-663	<i>Streptomyces capoamius</i>
21	ID.04-668	<i>Streptomyces roseoflavus</i>
22	ID.04-674	
23	ID.04-677	<i>Streptomyces peucetius</i>
24	ID.04-681	<i>Streptomyces capoamius</i>
25	P1 CCA9	
26	P3 CCA5	
27	P3 CCA6	
28	P3 CCA13	
29	P5 CCA2	
30	P5 CCA5	
31	P5 CCA7	
32	P6 CCA5	
33	CESE 01 RCT 1	
34	CLI 04 RC 1B	<i>Bacillus tequilensis</i>
35	TKA 01 RCC 4	
36	TTA 01 SDS 6	<i>Streptomyces luteovorticillatus</i> strain 173699
37	TTA 01 SDS 15	<i>Streptomyces seoulensis</i> strain NBRC 16668
38	TTA 01 SDST 2	
39	TTA 02 SDST 6	<i>Streptomyces spectabilis</i> (99%)
40	TTA 02 SDST 13	<i>Streptomyces mashuensis</i> (99%)
41	TCA 01 SDS 20	
42	TSA 01 SDS 1	<i>Streptomyces shandongensis</i> (83%)
43	TSA 02 SDS 2	<i>Kitasatospora putterlickiae</i> strain F18-98
44		<i>Streptomyces hygroscopicus</i> subsp. <i>Hygroscopicus</i> (99%)
	TSA 02 TC 4	
45	TSA 02 SDST 4	<i>Streptomyces fumigatiscleroticus</i> strain S6SS3
46	TSA 03 SDST 7	<i>Streptomyces anandii</i> strain NRRL B-3590
47	TSA 04 SDS 12	
48	MAUNA 5	

No	Isolate code	Name of isolates
49	MAUNA 6	
50	SA-15	
51	CLI04 RC-7	<i>Streptomyces polychromogenes</i>
52	CLI04 RC-2	<i>Actinoplanes brasiliensis</i>
53	CS 12 SDST-1	<i>Streptomyces thermodiastaticus</i>
54	Cwer03 E-2	<i>Ammycolatopsis saalfeldensis</i>
55	TCA01 SDS-13	<i>Streptomyces griseovorticillatus</i>
56	TCA01 SDS-14	<i>Streptomyces endus</i>
57	TCA01 SDS-17	<i>Nocardia arthritidis</i>
58	TKA01 SDS-8	<i>Streptomyces fradiae</i>
59	TSA02 SDS-12	<i>Streptomyces pulveraceus</i>
60	TTA 01 SDST-6	<i>Streptomyces sp</i>
61	TTA02 SDS-9	<i>Streptomyces puniscabiei</i>
62	TTA 02 SDS-13	<i>Streptomyces hygroscopicus</i> subsp. <i>Jinggangensis</i>
63	TTA 02 SDS-14	<i>Streptomyces yeochonensis</i> (84%)
64	TTA 02 SDST-2	<i>Streptomyces thermocarboxydovorans</i> strain AT52
65	TTA 02 SDST-18	<i>Streptomyces mashuensis</i> strain DSM 40221
66	GAL-1	
67	GAL-2	
68	GAL-3	
69	GAER-1	
70	GBL-4	
71	GCER-1	
72	GBSL-1	
73	GBSL-2	
74	GBSL-3	
75	GBSL-4	
76	GBSL-5	
77	GBSL-6	
78	GBSL-7	
79	GBSL-8	
80	GKRL-1	
81	GKRL-2	
82	GKRL-3	
83	GKRL-4	
84	GKRL-5	
85	GKRL-6	
86	GKRL-7	
87	GKRL-8	
88	GKRL-9	
89	GKRL-10	
90	GKRL-11	
91	GKRL-12	
92	GKRL-13	
93	ID.04-450	
94	ID.04-600	<i>Streptomyces peruviansis</i>
95	ID.04-686	<i>Streptomyces peucetius</i>
96	ID.04-687	<i>Streptomyces virginiae</i>
97	ID.04-691	<i>Streptomyces somaliensis</i>
98	ID.04-700	<i>Streptomyces galliaeus</i>
99	ID.04-766	<i>Streptomyces latentius</i>
100	ID.04-783	

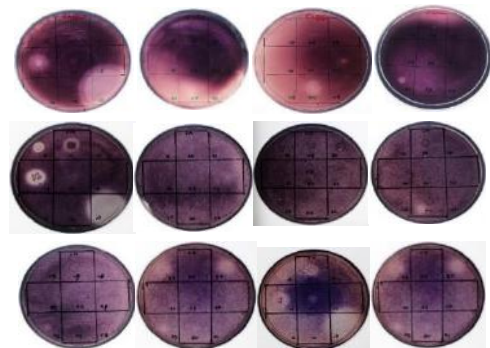
on CMC plate medium for cellulase



on Xylan plate medium for xylanase



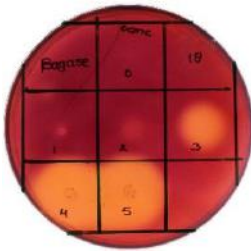
on LBG plate medium for mannase



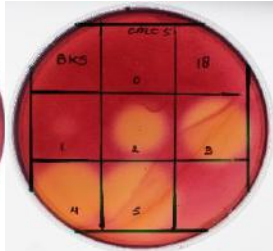
Many kinds of actinomycetes isolates show high activities of cellulase, xylanase and mannase

Production of Cellulase from Actinomycetes from many kinds of practical biomass

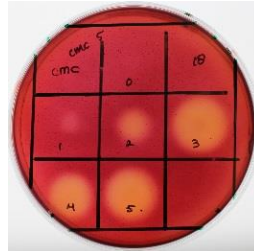
**Sugar Cane
Bagasse**



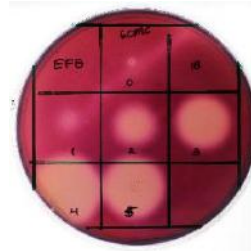
**Palm Kernel
Cake**



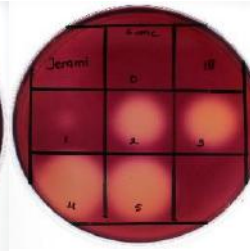
CMC



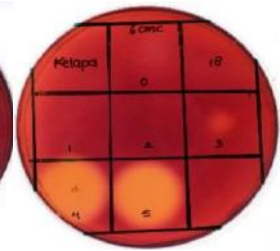
**Oil Palm Empty
Fruit Bunch**



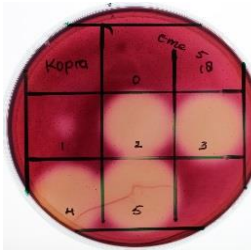
**Rice
Straw**



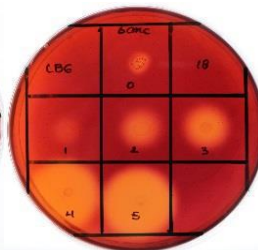
Coconut



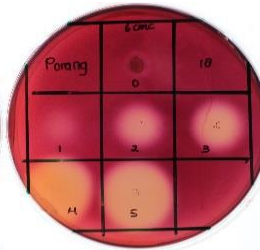
Kopra



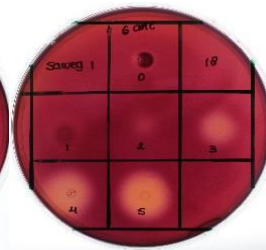
LBG



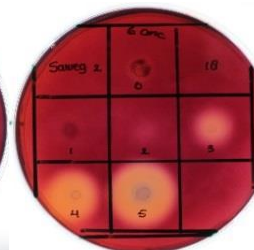
Porang



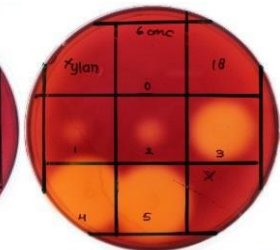
Suweg 1



Suweg 2

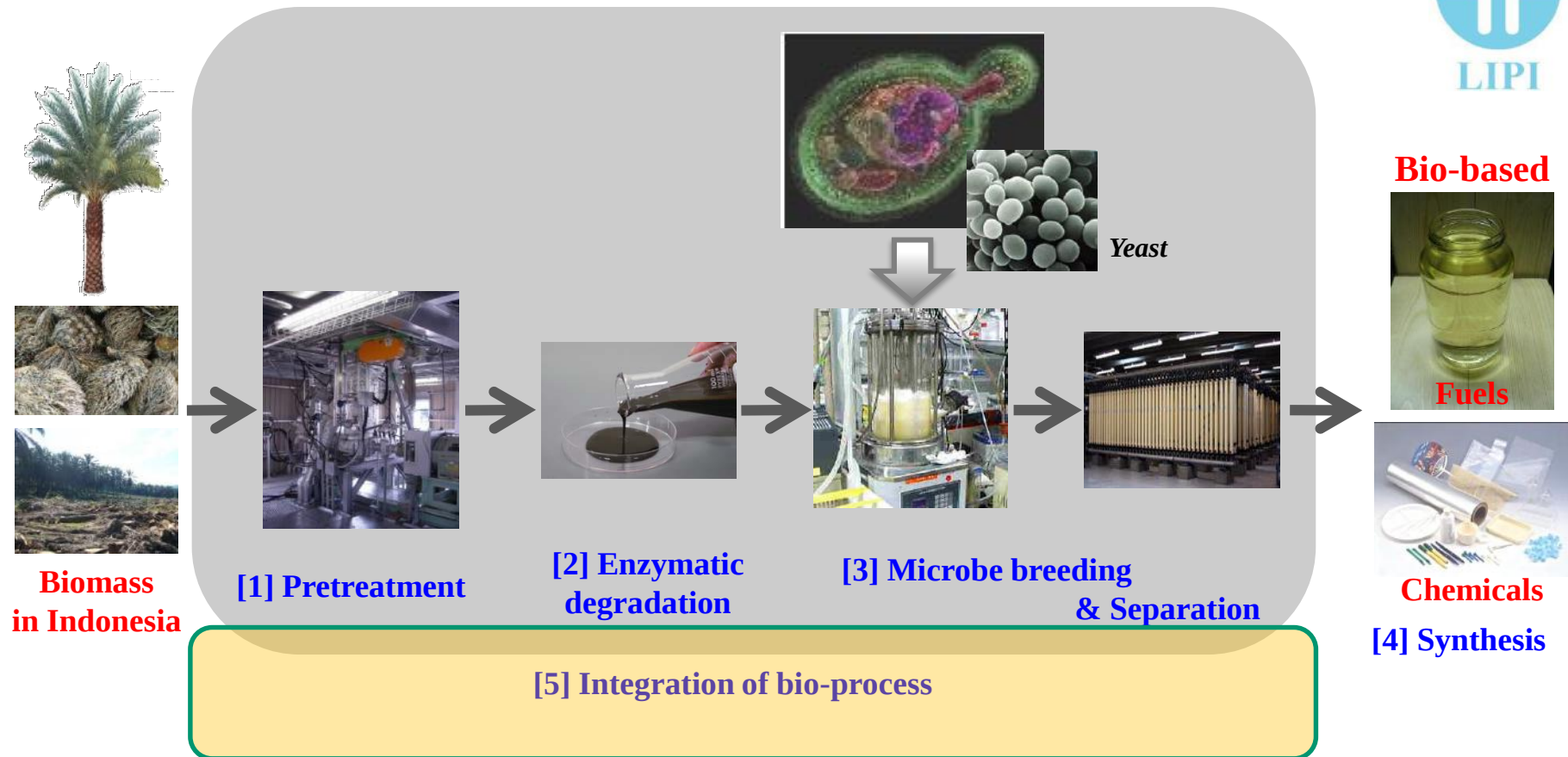


Xylan

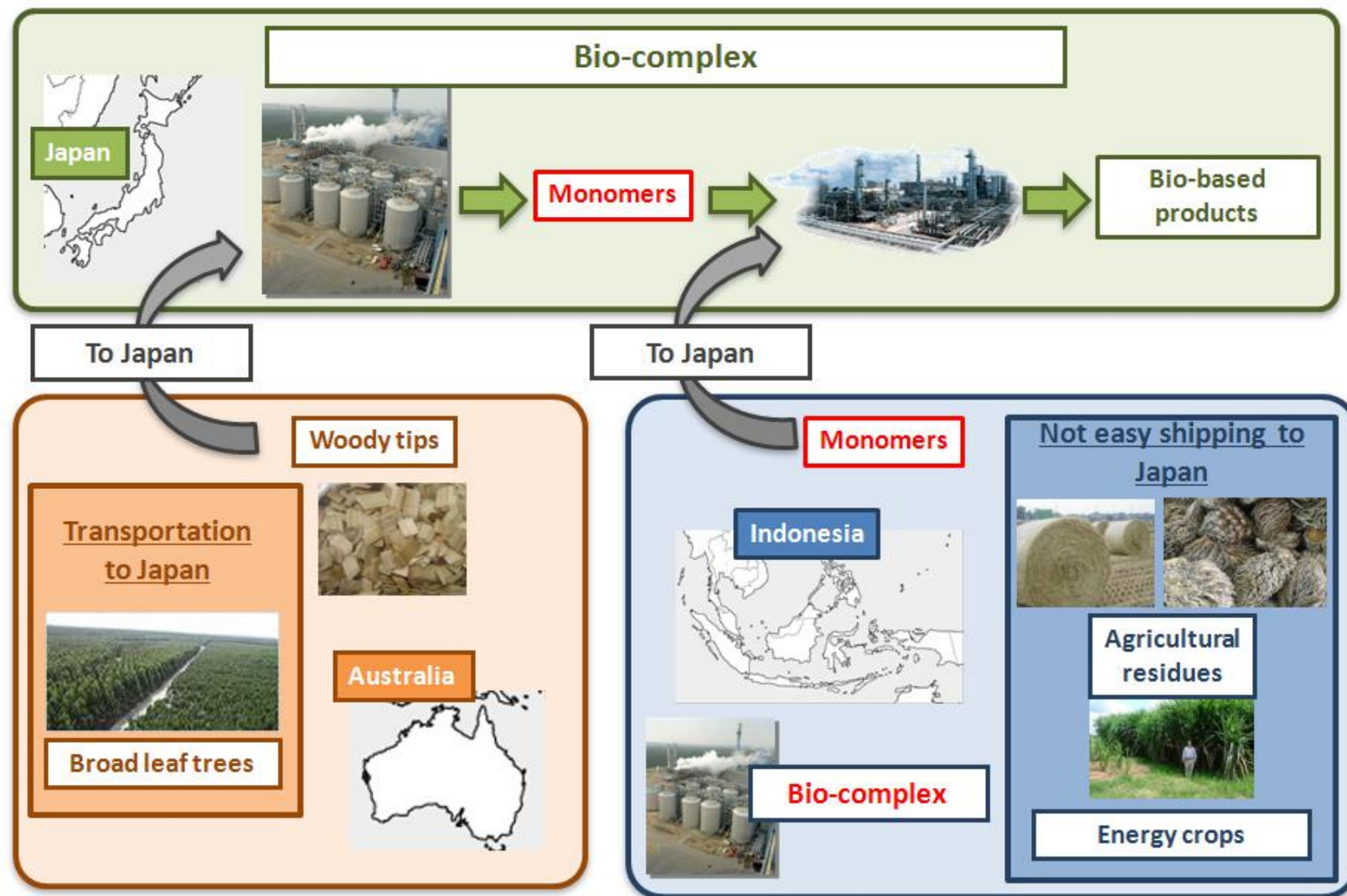


Isolate (No. 18) could produce Cellulase at high concentration and high enzyme activity from many kinds of practical biomass such as sugarcane bagasse, palm kernel cake, oilpalm empty fruit bunch, rice straw, etc, more better than that from CMC.

Five Key Technologies in SATREPS project



Establishment of Bio-Complex and biomass supply



White paper of biomass in Indonesia

ASSESSMENT OF PROSPECTS OF BIOMASS FOR BIOREFINERY INDUSTRI IN INDONESIA



EFB at an OP Mill



Bioethanol plant

By

**Manaek Simamora
Syahrizal Maulana
Syafrizal Maludin
Aris Yaman
Sasa Sofyan Munawar**

Business meeting between Japan and Indonesia



Process Development

Bench-scale plant in Kobe University



50 kg/h presser



100 kg/batch steamer



100 L
liquefaction reactor



50 L fermentation reactor



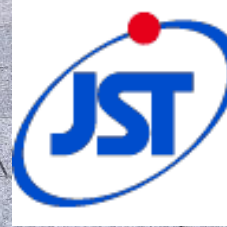
30 L fermentation reactor (x3)



Cell recycling system

January 2017

Acknowledgement to SATREPS member



Biomass complex strategy for chemical production

